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Expeditious synthesis of 5,6,7,8-tetrahydro-imidazo- [1,2-a]pyrimidin-2-ones and 3,4,6,7,8,9-hexahydro-pyrimido- $[1,2-a]$ pyrimidin-2-ones^{*}

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Abstract—A convenient synthesis of new 5,6,7,8-tetrahydro-imidazo[1,2-a]pyrimidin-2-ones and 3,4,6,7,8,9-hexahydro-pyrimido[1,2 a]pyrimidin-2-ones from the Baylis–Hillman adducts of acrylonitrile and their derivatives is described. A common strategy employed to achieve the syntheses of title compounds involved generation of diamines from different Baylis–Hillman derivatives followed by treatment with cyanogen bromide at reflux temperature to trigger a double intramolecular cyclization.

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1. Introduction

The versatility of Baylis–Hillman derivatives for the construction of diverse heterocyclic frameworks is stupendous. This is apparent by a recent surge in the number of publications describing synthetic methodologies toward this objective.[1](#page-6-0) One of our interests in this area concerns achieving practical synthesis of heterocyclic systems from Baylis– Hillman derivatives containing a nitrile group.^{[2](#page-7-0)} We have been especially interested in the synthesis of ring systems incorporating guanidine functionality. In this context, recently, we have reported the solid-phase synthesis of annulated pyrimidinones from the Baylis–Hillman adducts via Michael addition of alkanediamines followed by cyanogen bromidemediated cyclization of the diamine and finally basepromoted cyclative cleavage.[3](#page-7-0) The adoption of this strategy on the solid-phase chemistry was a direct outcome of our unsuccessful attempt to carry out the Michael addition of alkanediamines onto the Baylis–Hillman acetates in solutionphase as it invariably led to the formation of the polymeric products. Intriguingly, imidazo-pyrimidinone system is not very well represented in the literature though some of the derivatives bearing this moiety have been reported to display significant bioactivities.^{[4](#page-7-0)} With the aim to diversify the synthesis of annulated [1,2-a]pyrimidin-2-ones in solution-phase, we decided to utilize the derivatives of Baylis–Hillman adduct of acrylonitrile. Conceptually, the Michael addition of an amino alkyl ester on the Baylis–Hillman adduct of acrylonitrile followed by reduction of the nitrile function would lead to a diamino system, which may be cyclized with cyanogen bromide to yield the annulated $[1,2-a]$ pyrimidine-2ones. On the other hand, the alkylation of a primary allyl amine, 5 generated from the acetyl derivative of Baylis– Hillman adducts, with alkyl haloacetate should lead to an allyl amino alkyl ester. Subsequently, reducing the nitrile to amine would also furnish a diamino framework, which should undergo a double intramolecular cyclization by reaction with cyanogen bromide to afford an annulated [1,2-a]pyrimidin-2 one system. Alternatively, acetates can be subjected to Michael addition with amino alkyl ester followed by reduction of nitrile group and successive cascade cyclizations with cyanogen bromide. It was reasoned that this strategy might be extended to the synthesis of pyrimido $[1,2-a]$ pyrimidin-2-ones. In principle, primary allyl amine afforded from the acetyl derivative of Baylis–Hillman adducts of acrylonitrile would undergo S_N2 reaction with the acetyl derivative of Baylis– Hillman adduct of acrylate to yield a product, which may serve as substrate for the synthesis of the desired heterocyclic system. Herein, we describe the details of our successful attempts toward expeditious synthesis of a variety of annulated [1,2-a]pyrimidin-2-ones involving these approaches.

2. Results and discussion

2.1. Synthesis of 6-(hydroxy-substituted phenyl-methyl)- 5,6,7,8-tetrahydro-imidazo[1,2-a]pyrimidin-2-ones

We initiated our studies from the Baylis–Hillman adduct of acrylonitrile 1a, which was easily obtained by following standard procedure.^{[2a](#page-7-0)} Subsequent reaction of glycine ester

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with **1a** at reflux temperature for 12 h gave the corresponding product 2a as diastereoisomeric mixture (1:1) in excellent yield (Scheme 1). Hydrogenating 2a in the presence of Raney-Ni for 3 h afforded the desired diamine 3a. The diamine 3a being unstable in nature was immediately subjected to reaction with cyanogen bromide in absolute ethanol at reflux temperature to yield the substituted imidazo[1,2 a]pyrimidine-2-one 4a as hydrobromide salt in good yield (Scheme 1). The generality of this reaction protocol was evident from the synthesis of several analogs 4b–g. The spectroscopic data especially the UV spectra of compounds 4a–g supported the assigned structure to be the 5,6,7,8-tetrahydro-imidazo $[1,2-a]$ pyrimidine-2-ones.^{[4a](#page-7-0)}

2.2. Synthesis of 6-benzyl-5,6,7,8-tetrahydro-imidazo[1,2-a]pyrimidin-2-ones and 6-Methyl-5-phenyl-5,6,7,8-tetrahydro-imidazo[1,2-a]pyrimidin-2-ones

In order to increase the scope of this strategy, it was decided to carry out similar reactions with the acetyl derivative of the Baylis–Hillman adducts of acrylonitrile. In view of our earlier observation that during the reaction of the acetyl derivatives of the Baylis–Hillman adducts large excess of primary amine has to be used, 5 we decided to avoid the reaction of glycine ester directly with 5. Therefore, the allyl amine 6,

generated via a reported procedure,^{[5](#page-7-0)} was treated with ethyl iodoacetate in THF in the presence of K_2CO_3 for 1.5 h to yield a mixture of two products (Scheme 2). These products were isolated in 4 and 58% yields and were established to be 7 and 8, respectively (Scheme 2). Alternatively, the treatment of amine 6 with ethyl bromoacetate yielded the desired secondary amine 8 in 63% yield with only traces of 7, though the reaction was completed in 15 h. Hydrogenating the allyl amine 8 with Raney-Ni gave the diamino derivative 9 in good yield. The product 9 was immediately treated with cyanogen bromide in absolute alcohol at reflux temperature to furnish the product 10 (Scheme 2). Encouraged with these results, we decided to extend the utility of this method to the synthesis of substituted imidazo $[1,2-a]$ pyrimidin-2-ones from the substrate afforded by the S_N2 reaction of glycine ester and the acetyl derivative of Baylis–Hillman adduct. Therefore, the acetates 5c,g were treated with glycine ester in the presence of DABCO in a THF/water system $(1:1, v/v)$ to furnish corresponding products $11c, g$ (Scheme 3). Reduction of the nitrile group with Raney-Ni in 11c,g led to the isolation of diamines 12c,g, respectively. Treatment of diamines 12c,g with cyanogen bromide in absolute ethanol at reflux temperature afforded the desired 5,6,7,8-tetrahydro-imidazo[1,2-a]pyrimidine-2-one 13c,g as diastereoisomeric mixture in 1:1 ratio.

Compd. R

2.3. Synthesis of 7-substituted phenyl-methyl-3-methyl-4-substituted phenyl-3,4,6,7,8,9-hexahydro-pyrimido[1,2-a]pyrimidin-2-ones

Buoyed by the success of our strategy for the generation of a variety of imidazo[1,2-a]pyrimidinone system, we decided to investigate the synthesis of pyrimido $[1,2-a]$ pyrimidin-2ones. In principle, S_N2 reaction between the primary allyl amine 6 and the acetyl derivative of Baylis–Hillman adduct of acrylate would yield a suitable starting substrate for the desired motif. Hence, treatment of compounds 6a–c with 14 and 15 in the presence DABCO in a THF/water system $(1:1, v/v)$ furnished the products $16-19$ in excellent yields (Scheme 4). Hydrogenation of 16–19 with Raney-Ni and subsequent treatment of the diamine 20–23 with cyanogen bromide in absolute ethanol gave the desired 3,4,6,7,8,9 hexahydro-pyrimido[1,2-a]pyrimidin-2-ones 24–27 in moderate yields as diastereoisomeric mixture in 1:1 ratio.

3. Conclusions

In summary, we have demonstrated convenient and practical strategies for the synthesis of differently substituted new annulated 5,6,7,8-tetrahydro-imidazo[1,2-a] pyrimidine-2 ones and $3,4,6,7,8,9$ -hexahydro-pyrimido $[1,2-a]$ pyrimidin-2-ones from the derivatives of Baylis–Hillman adducts of acrylonitrile.

4. Experimental

4.1. General

Melting points are uncorrected and were determined in capillary tubes on an apparatus containing silicon oil. IR spectra were recorded using Perkin–Elmer Spectrum RX I FTIR spectrophotometer. ¹H NMR and ¹³C NMR spectra were recorded either on a Bruker DPX-200 FT or Bruker Avance DRX-300 spectrometers, using TMS as an internal standard (chemical shifts in δ values, J in Hz). The ESMS were recorded on MICROMASS LC–MS system and FABMS were recorded on JEOL/SX-102 spectrometer. Elemental analyses were performed on Carlo Erba 108 or Elementar Vario EL III microanalyzer. The HRMS spectra were recorded as EI-HRMS. The HPLCs were performed on RP C-18 column (250×4.5 mm, $5 \mu m$) using a gradient of 10– 100% methanol containing 0.1% TFA and water in 25 min at a flow rate of 1 mL/min on Agilent 1100. The silica gel used for column chromatography of compounds 4a–g, 10, 13c,g, and 24–27 was deactivated prior to use by adding 12 mL of water per 100 g of silica gel. Compounds 2a–g, 4a–g, 13c,g, and 24–27 were obtained as diastereoisomeric mixture in $1:1$ ratio as evident from the 1 H NMR of the crude products.

4.2. General procedure for the preparation of 2a–g

To a stirred solution of appropriate compound from 1a–g (3.16 mmol) in MeOH (5 mL) were added Et₃N (6.3 mmol, 0.65 mL) and glycine methyl ester hydrochloride (4.74 mmol, 0.6 g). The reaction mixture was heated at reflux temperature till the completion of reaction (monitored by TLC, ca. 12 h). Thereafter, methanol was removed in vacuo and the residue was extracted with EtOAc $(3\times25 \text{ mL})$ and water (40 mL). The organic layers were combined, dried over $Na₂SO₄$, and concentrated to afford crude products, which were purified via silica gel column chromatography using hexane/EtOAc (60:40, v/v) to yield the pure products.

4.2.1. (2-Cyano-3-hydroxy-3-phenyl-propylamino)-acetic acid methyl ester (2a). Yield: 92% as colorless oil; R_f =0.4 (hexane/EtOAc, 60:40); v_{max} (neat) 1739 (CO₂Me), 2245 (CN), 3331 (NH, OH) cm⁻¹; ¹H NMR (200 MHz, CDCl₃) δ 2.71 (br s, 2H, 2×NH), 2.92–3.10 (m, 4H, $2 \times CH_2NH$, 3.19–3.25 (m, 2H, 2 \times CH), 3.50 (s, 2H, CH₂CO), 3.53 (s, 2H, CH₂CO), 3.75 (s, 6H, 2×OCH₃), 5.07 (d, 1H, $J=3.0$ Hz, CHOH), 5.08 (d, 1H, $J=3.0$ Hz, CHOH), $7.33-7.47$ (m, 10H, $2\times5ArH$); ¹³C NMR $(75 \text{ MHz}, \text{CDCl}_3)$ δ 43.5, 49.5, 50.3, 52.1, 53.1, 53.6, 71.9, 73.3, 74.1, 120.1, 121.1, 121.4, 127.7, 127.8, 128.0, 128.2, 129.8, 129.4, 130.1, 142.6, 142.8, 143.1, 166.0, 174.1; mass $(FAB+)$ $m/z=249$ $(M^+ + 1)$; Anal. Calcd for $C_{13}H_{16}N_2O_3$: C, 62.89; H, 6.50; N, 11.28. Found: C, 63.13; H, 6.28; N, 11.35.

4.2.2. (2-Cyano-3-hydroxy-3-naphthalen-2-propylamino)-acetic acid methyl ester (2b). Yield: 83% as colorless oil; R_f =0.6 (hexane/EtOAc, 60:40); v_{max} (neat) 1739 $(CO₂Me)$, 2244 (CN), 3451 (NH, OH) cm⁻¹; ¹H NMR $(300 \text{ MHz}, \text{ CDCl}_3)$ δ 2.92–3.06 (m, 4H, 2×CH₂NH), 3.16–3.23 (m, 2H, 2 \times CH), 3.42–3.50 (m, 4H, 2 \times CH₂CO), 3.78 (s, 6H, $2 \times OCH_3$), 5.23 (d, 1H, $J=2.8$ Hz, CHOH), 5.29 (d, 1H, $J=2.8$ Hz, CHOH), 7.49–7.52 (m, 6H, 2×3 ArH), 7.81–7.93 (m, 8H, 2×4 ArH); ¹³C NMR (50 MHz, CDCl3) d 11.3, 14.6, 40.1, 41.3, 48.5, 49.7, 50.5, 50.7, 52.5, 60.9, 61.6, 73.7, 74.3, 76.9, 77.5, 78.2, 119.1, 119.6, 124.0, 124.2, 125.6, 126.7, 126.8, 128.6, 128.9, 133.6, 133.7, 138.3, 172.6; mass (ES+) $m/z = 299$ (M⁺+1);

Anal. Calcd for $C_{17}H_{18}N_2O_3$: C, 68.44; H, 6.08; N, 9.39. Found: C, 68.29; H, 5.90; N, 9.57.

4.2.3. (2-Cyano-2-hydroxy-3-p-tolyl-propylamino)-acetic acid methyl ester (2c). Yield: 92% as colorless oil; R_f =0.5 (hexane/EtOAc, 60:40); v_{max} (neat) 1739 (CO₂Me), 2245 (CN) , 3322 (NH, OH) cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 2.36 (s, 6H, 2×CH₃), 2.56 (br s, 2H, 2×NH), 2.91–3.06 $(m, 6H, 2 \times CH \text{ and } 2 \times CH_2NH), 3.48-3.50 \text{ (m, 4H)}$ $2 \times CH_2CO$), 3.75 (s, 6H, $2 \times OCH_3$), 5.00 (d, 1H, $J=4.7$ Hz, CHOH), 5.03 (d, 1H, $J=4.6$ Hz, CHOH), 7.20 (d, 4H, $J=8.0$ Hz, $2\times$ ArH), 7.33 (d, 4H, $J=8.0$ Hz, 2×4 ArH); ¹³C NMR (75 MHz, CDCl₃) δ 12.9, 19.9, 38.6, 39.7, 47.0, 48.0, 48.8, 49.0, 50.8, 59.2, 72.1, 72.5, 117.4, 117.8, 124.5, 124.9, 128.0, 136.1, 136.9, 137.1, 170.9, 171.0; mass (ES+) $m/z = 263$ (M⁺+1); Anal. Calcd for $C_{14}H_{18}N_2O_3$: C, 64.10; H, 6.92; N, 10.68. Found: C, 63.82; H, 7.05; N, 10.51.

4.2.4. [3-(2-Bromo-phenyl)-2-cyano-3-hydroxy-propylamino]-acetic acid methyl ester (2d). Yield: 93% as colorless oil; R_f =0.5 (hexane/EtOAc, 60:40); ν_{max} (neat) 1717 (CO_2Me) , 2200 (CN), 3463 (NH, OH) cm⁻¹; ¹H NMR (200 MHz, CDCl₃) δ 2.93–3.22 (m, 4H, 2×CH₂NH), $3.23-3.32$ (m, 2H, 2 \times CH), 3.41 (s, 2H, CH₂CO), 3.60 (d, 2H, J=0.5 Hz, CH₂CO), 3.75 (s, 3H, OCH₃), 3.77 (s, 3H, OCH₃), 5.37 (d, 1H, $J=1.4$ Hz, CHOH), 5.48 (d, 1H, J=3.0 Hz, CHOH), 7.20-7.25 (m, 2H, ArH), 7.39-7.60 (m, 4H, ArH), 7.82–7.89 (m, 2H, ArH); 13C NMR $(75 \text{ MHz}, \text{CDCl}_3)$ δ 35.5, 37.0, 45.9, 48.8, 49.0, 50.9, 71.3, 71.9, 116.6, 117.9, 119.8, 120.7, 126.6, 126.8, 127.1, 127.2, 128.5, 128.7, 131.2, 131.8, 137.8, 138.0, 170.7, 170.8; mass (ES+) $m/z = 328$ (M⁺), 330 (M⁺+2); Anal. Calcd for $C_{13}H_{15}BrN_2O_3$: C, 47.72; H, 4.62; N, 8.56. Found: C, 47.97; H, 4.78; N, 8.22.

4.2.5. [3-(2-Chloro-phenyl)-2-cyano-2-hydroxy-3-propylamino]-acetic acid methyl ester (2e). Yield: 89% as colorless oil; R_f =0.6 (hexane/EtOAc, 60:40); v_{max} (neat) 1740 (CO_2Me) , 3430 (NH, OH) cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 2.93–3.20 (m, 4H, 2×CH₂NH), 3.37–3.44 (m, 2H, 2×CH), 3.50 (s, 2H, CH₂CO), 3.60 (s, 2H, CH₂CO), 3.75 (s, 3H, OCH3), 3.78 (s, 3H, OCH3), 5.45 (d, 1H, $J=4.2$ Hz, CHOH), 5.53 (d, 2H, $J=4.3$ Hz, CHOH), 7.32– 7.42 (m, 5H, ArH), 7.62–7.66 (m, 1H, ArH), 7.83–7.87 (m, 2H, ArH); ¹³C NMR (75 MHz, CDCl₃) δ 36.1, 37.3, 38.6, 38.9, 45.9, 48.6, 49.0, 50.9, 68.5, 69.2, 116.8, 118.1, 126.0, 126.1, 126.9, 127.9, 128.1, 128.3, 128.5, 129.6, 130.6, 136.7, 170.9; mass (ES+) $m/z=283$ (M⁺+1), 285 $(M^+ + 3)$; Anal. Calcd for C₁₃H₁₅ClN₂O₃: C, 55.23; H, 5.35; N, 9.91. Found: C, 55.01; H, 5.54; N, 10.11.

4.2.6. [2-Cyano-3-(2-fluoro-phenyl)-3-hydroxy-propylamino]-acetic acid methyl ester (2f). Yield: 79% as colorless oil; R_f =0.5 (hexane/EtOAc, 60:40); ν_{max} (neat) 1734 $(CO₂Me)$, 2233 (CN), 3437 (NH, OH) cm⁻¹; ¹H NMR $(300 \text{ MHz}, \text{ CDCl}_3)$ δ 2.28 (br s, 2H, 2×NH), 2.95–3.14 $(m, 6H, 2 \times CH_2NH)$ and CH, 3.49 (s, 2H, CH₂CO), 3.57 (s, 2H, CH2CO), 3.74 (s, 3H, OCH3), 3.76 (s, 3H, OCH3), 5.37 (d, 1H, $J=2.6$ Hz, CHOH), 5.38 (d, 1H, $J=2.6$ Hz, CHOH), 7.01–7.11 (m, 2H, ArH), 7.21–7.32 (m, 4H, ArH), 7.58–7.59 (m, 2H, ArH); ¹³C NMR (75 MHz, CDCl3) d 35.6, 37.1, 45.8, 48.7, 48.9, 50.8, 50.9, 71.1, 71.8, 116.7, 118.0, 119.8, 120.7, 126.6, 126.8, 127.1, 127.2, 128.5, 127.7, 131.2, 131.8, 137.9, 138.0, 170.7, 170.9; mass (ES+) $m/z=267$ (M⁺+1); Anal. Calcd for $C_{13}H_{15}FN_2O_3 \cdot H_2O$: C, 54.92; H, 6.03; N, 9.85. Found, C, 55.12; H, 5.88; N, 10.05.

4.2.7. [2-Cyano-3-(2,4-dichloro-phenyl)-3-hydroxy-3 propylamino]-acetic acid methyl ester (2g). Yield: 81% as colorless oil; $R_f=0.7$ (hexane/EtOAc, 60:40); v_{max} (neat) 1740 (CO₂Me), 3430 (NH, OH) cm⁻¹; ¹H NMR $(300 \text{ MHz}, \text{ CDCl}_3)$ δ 3.08–3.20 (m, 4H, 2×CH₂NH), $3.37-3.50$ (m, 6H, $2 \times CH$ and CH_2CO), 3.75 (s, 3H, OCH₃), 3.78 (s, 3H, OCH₃), 5.42 (d, 1H, J=4.6 Hz, CHOH), 5.50 (d, 2H, $J=4.6$ Hz, CHOH), 7.36-7.40 (m, 4H, ArH), 7.56–7.61 (m, 1H, ArH), 7.78–7.82 (m, 1H, ArH); ¹³C NMR (75 MHz, CDCl₃) δ 35.6, 36.9, 46.2, 48.9, 51.0, 60.1, 69.3, 70.0, 116.6, 126.2, 126.9, 127.9, 128.1, 128.3, 128.6, 129.6, 136.3, 170.7, 170.9; mass (ES⁺) $m/z = 317$ (M⁺+1), 319 (M⁺+3); Anal. Calcd for $C_{13}H_{14}Cl_2N_2O_3$: C, 49.23; H, 4.45; N, 8.83. Found: C, 49.28; H, 4.20; N, 8.86.

4.3. General procedure for the preparation of 3a–g, 9, 12c,g, and 20–23

A mixture of appropriate compound from 2a–g, 8 or 11c,g, 16–19 (2.0 mmol) and Raney-Ni (0.1 g wet) in methanol (10 mL) was subjected to hydrogenation at 40 psi in the Parr assembly for 3 h. The catalyst was filtered through a pad of Celite and the filtrate was concentrated. The residue was used directly for further reactions.

4.4. General procedure for the preparation of 4a–g, 10, 13c,g, and 24–27

To a stirred solution of appropriate diamines from 3a–g, 9, 12c,g or $20-23$ (1.98 mmol) in absolute EtOH (15 mL) was added CNBr (2.97 mmol, 0.315 g) and the reaction mixture was heated at reflux temperature for 8 h (36–48 h for 20–23). Ethanol was removed from the mixture and the residue was subjected to silica gel column chromatography using (CHCl₃/MeOH, 90:10, v/v) as the eluant to furnish the pure products as hydrobromide salts.

4.4.1. 6-(Hydroxy-phenyl-methyl)-5,6,7,8-tetrahydroimidazo $[1,2-a]$ pyrimidin-2-one (4a). Yield: 57% as a brown oil; R_f =0.3 (CHCl₃/MeOH, 90:10); ν_{max} (neat) 1658 (CONH), 3370 (NH, OH) cm^{-1} ; ¹H NMR $(200 \text{ MHz}, \text{ DMSO-}d_6)$ δ 2.32–2.49 (m, 2H, 2×CH), 2.76– 2.89 (m, 4H, $2 \times CH_2N$), 3.11–3.23 (m, 4H, $2 \times CH_2NH$), 3.56 (s, 4H, $2 \times CH_2CO$), 5.03 (d, 1H, $J=3.6$ Hz, CHOH), 5.11 (d, 1H, $J=3.5$ Hz, CHOH), 7.48–7.61 (m, 4H, ArH), 7.88–7.93 (m, 6H, ArH); ¹³C NMR (50 MHz, DMSO- d_6) d 37.1, 37.5, 42.9, 43.5, 49.9, 51.6, 69.6, 72.6, 125.3, 126.7, 126.9, 127.3, 127.8, 128.4, 128.6, 143.5, 144.2, 158.5, 158.7, 172.2, 172.5; mass (ES+) $m/z = 246$ (M⁺+1); λ_{max} =226 nm; HR-EIMS Calcd for C₁₃H₁₅N₃O₂: 245.1164, found: 245.1166.

4.4.2. 6-(Hydroxy-naphthalen-2-yl-methyl)-5,6,7,8-tetrahydro-imidazo[1,2-a]pyrimidin-2-one (4b). Yield: 62% as a brown oil; $R_f = 0.5$ (CHCl₃/MeOH, 90:10); v_{max} (neat) 1661 $(CONH)$, 3427 (NH, OH) cm⁻¹; ¹H NMR (200 MHz,

DMSO- d_6) δ 2.26–2.28 (m, 2H, 2×CH), 3.05–3.19 (m, 4H, $2 \times CH_2N$), 3.46–3.74 (m, 4H, $2 \times CH_2NH$), 3.75 (s, 4H, $2 \times CH_2CO$), 4.87 (d, 1H, J=4.2 Hz, CHOH), 4.92 (d, 1H, J=4.2 Hz, CHOH), 7.42–7.52 (m, 6H, ArH), 7.75–7.86 (m, 8H, ArH); ¹³C NMR (50 MHz, DMSO- d_6) δ 37.3, 37.8, 41.4, 41.9, 42.5, 43.0, 51.6, 52.0, 73.3, 73.9, 126.4, 127.2, 129.1, 129.7, 130.1, 130.7, 131.5, 131.8, 132.0, 133.2, 133.4, 138.3, 138.9, 146.7, 147.1, 158.0, 158.7, 177.6, 178.3; mass (ES+) $mlz=296$ (M⁺+1); $\lambda_{\text{max}}=224$ nm; HR-EIMS Calcd for $C_{17}H_{17}N_3O_2$: 295.1321, found: 295.1325.

4.4.3. 6-(Hydroxy-p-tolyl-methyl)-5,6,7,8-tetrahydroimidazo $[1,2-a]$ pyrimidin-2-one (4c). Yield: 89% as a colorless oil; $R_f=0.4$ (CHCl₃/MeOH, 90:10); ν_{max} (neat) 1662 $(CONH)$, 3409 (NH, OH) cm⁻¹; ¹H NMR (300 MHz, DMSO- d_6) δ 2.30 (s, 6H, 2×CH₃), 2.80–2.83 (m, 2H, $2 \times CH$), 3.06–3.11 (m, 4H, $2 \times CH_2N$), 3.27–3.43 (m, 4H, $2 \times CH_2NH$, 3.83 (s, 4H, $2 \times CH_2CO$), 4.64 (d, 1H, J= 4.4 Hz, CHOH), 4.69 (d, 1H, J=4.4 Hz, CHOH), 7.18 (d, 4H, J=8.0 Hz, ArH), 7.27 (d, 4H, J=8.0 Hz, ArH), 7.78 (br s, 2H, 2×NH), 10.78 (s, 2H, 2×OH); 13 C NMR $(75 \text{ MHz}, \text{ DMSO-}d_6)$ δ 21.2, 24.6, 24.7, 43.5, 43.9, 45.3, 45.8, 47.0, 47.8, 55.2, 56.3, 63.7, 76.5, 77.1, 130.3, 131.4, 132.7, 132.9, 140.5, 142.1, 144.0, 161.7, 161.9, 173.9, 175.8; mass (ES+) $m/z = 260$ (M⁺+1); $\lambda_{\text{max}} = 220$ nm; HR-EIMS Calcd for $C_{14}H_{17}N_3O_2$: 259.1321, found: 259.1349.

4.4.4. 6-[(2-Bromo-phenyl)-hydroxy-methyl]-5,6,7,8 tetrahydro-imidazo[1,2-a]pyrimidin-2-one (4d). Yield: 66% as colorless oil; R_f =0.4 (CHCl₃/MeOH, 90:10); ν_{max} $(neat)$ 1658 (CONH), 3370 (NH, OH) cm⁻¹; ¹H NMR $(300 \text{ MHz}, \text{ DMSO-}d_6)$ δ 2.30–2.36 (m, 2H, 2×CH), 3.02– 3.29 (m, 8H, $2 \times CH_2N$ and $2 \times CH_2NH$), 3.67 (s, 4H, $2 \times CH_2CO$), 4.87 (d, 1H, J=4.4 Hz, CHOH), 4.89 (d, 1H, J¼4.4 Hz, CHOH), 7.22–7.27 (m, 2H, ArH), 7.42–7.46 (m, 2H, ArH), 7.54–7.58 (m, 4H, ArH); 13C NMR $(50 \text{ MHz}, \text{ DMSO-}d_6)$ δ 37.6, 37.9, 39.5, 40.7, 41.8, 42.1, 52.0, 52.6, 71.2, 71.9, 121.6, 125.4, 126.8, 127.6, 128.2, 129.0, 129.8, 133.0, 142.2, 158.0, 172.1, 172.6; mass (ES+) $m/z = 324$ (M⁺+1); $\lambda_{\text{max}} = 226$ nm; HR-EIMS Calcd for $C_{13}H_{14}BrN_3O_2$: 323.0269, found: 323.0265.

4.4.5. 6-[(2-Chloro-phenyl)-hydroxy-methyl]-5,6,7,8 tetrahydro-imidazo[1,2-a]pyrimidin-2-one (4e). Yield: 83% as white solid; mp 272–274 °C; $R_f=0.5$ (CHCl₃/MeOH, 90:10); v_{max} (KBr) 1648 (CONH), 3415 (NH, OH) cm⁻¹;
¹H NMR (300 MHz, DMSO-d), δ 232-234 (m) 2H ¹H NMR (300 MHz, DMSO- d_6) δ 2.32–2.34 (m, 2H, $2 \times$ CH), 2.83–2.97 (m, 4H, $2 \times$ CH₂N), 3.25–3.32 (m, 4H, $2 \times CH_2NH$, 4.01 (s, 4H, $2 \times CH_2CO$), 5.05 (d, 1H, J= 4.5 Hz, CHOH), 5.09 (d, 1H, $J=4.5$ Hz, CHOH), 7.16– 7.31 (m, 6H, ArH), 7.58–7.60 (m, 2H, ArH), 7.87 (br s, 2H, $2\times$ NH), 10.89 (br s, 2H, $2\times$ OH); ¹³C NMR (50 MHz, DMSO-d₆) δ 37.6, 38.2, 40.2, 40.9, 41.7, 42.5, 51.4, 51.9, 69.0, 69.8, 127.7, 128.7, 129.0, 129.4, 129.7, 130.1, 131.2, 133.9, 140.8, 158.0, 158.6, 172.1, 172.9; mass (ES+) $m/z=280$ (M⁺+1); $\lambda_{\text{max}}=222$ nm; HR-EIMS Calcd for $C_{13}H_{14}CIN_3O_2$: 279.0775, found: 279.0778.

4.4.6. 6-[(2-Fluoro-phenyl)-hydroxy-methyl]-5,6,7,8-tetrahydro-imidazo[1,2-a]pyrimidin-2-one (4f). Yield: 74% as colorless oil; R_f =0.5 (CHCl₃/MeOH, 90:10); v_{max} (neat): 1676 (CONH), 3436 (NH, OH) cm⁻¹; ¹H NMR

 $(300 \text{ MHz}, \text{ DMSO-}d_6)$ δ 2.98–3.16 (m, 6H, 2×CH and $2 \times CH_2N$), 3.51–3.62 (m, 4H, $2 \times CH_2NH$), 3.77 (s, 4H, $2 \times CH_2CO$), 5.42 (d, 1H, J=4.5 Hz, CHOH), 5.45 (d, 1H, J=4.5 Hz, CHOH), 7.05-7.12 (m, 2H, ArH), 7.23-7.36 (m, 4H, ArH), 7.58–7.59 (m, 2H, ArH); 13C NMR $(50 \text{ MHz}, \text{ DMSO-}d_6)$ δ 38.5, 38.8, 41.7, 41.9, 42.3, 43.0, 51.6, 52.1, 73.0, 73.5, 126.7, 127.0, 127.3, 128.6, 129.1, 129.4, 130.0, 136.9, 140.4, 158.1, 158.7, 172.2, 173.1; mass (ES+) $m/z = 264$ (M⁺+1); $\lambda_{\text{max}} = 226$ nm; HR-EIMS Calcd for $C_{13}H_{14}FN_3O_2$: 263.1070, found: 263.1074.

4.4.7. 6-[(2,4-Dichloro-phenyl)-hydroxy-methyl]-5,6,7,8 tetrahydro-imidazo $[1,2-a]$ pyrimidin-2-one $(4g)$. Yield: 88% as a white solid; mp 248–250 °C; $R_f=0.6$ (CHCl₃/ MeOH, 90:10); v_{max} (KBr) 1662 (CONH), 3367 (NH, OH) cm⁻¹; ¹H NMR (300 MHz, DMSO- d_6) δ 2.24–2.26 (m, 2H, $2 \times$ CH), 2.76–2.79 (m, 4H, $2 \times$ CH₂N), 3.35–3.42 (m, 4H, $2 \times CH_2NH$), 3.88 (s, 4H, $2 \times CH_2CO$), 4.99 (d, 1H, J=3.9 Hz, CHOH), 5.01 (d, 1H, J=3.9 Hz, CHOH), 6.10 (s, 2H, 2-NH), 7.50–7.53 (m, 2H, ArH), 7.60–7.69 (m, 4H, ArH), 10.86 (s, 2H, 2×OH); ¹³C NMR (75 MHz, DMSO- d_6) δ 36.5, 36.8, 40.6, 41.9, 42.6, 50.7, 51.4, 67.6, 69.1, 126.7, 127.6, 128.0, 128.7, 129.0, 129.8, 131.0, 138.9, 156.8, 158.0, 170.9, 172.1; mass (ES+) $m/z=314$ (M⁺+1); $\lambda_{\text{max}}=230 \text{ nm}$; HR-EIMS Calcd for $C_{13}H_{12}Cl_2N_3O_2$: 313.0385, found: 313.0389.

4.4.8. 6-Benzyl-5,6,7,8-tetrahydro-imidazo[1,2-a]pyrimi**din-2-one (10).** Yield: 48% as a brown oil; $R_f=0.4$ (CHCl₃/ MeOH, 90:10); v_{max} (neat) 1660 (CONH), 3362 (NH) cm⁻¹;
¹H NMR (200 MHz, CDCL) δ 2 20-2 41 (m, 1H, CH), 2.67-¹H NMR (200 MHz, CDCl₃) δ 2.20–2.41 (m, 1H, CH), 2.67– 2.71 (m, 2H, CH₂N), 3.12–3.45 (m, 4H, ArCH₂ and CH₂NH), 3.88 (s, 2H, CH₂CO), 6.52 (br s, 1H, NH), 7.16– 7.38 (m, 5H, ArH); ¹³C NMR (50 MHz, DMSO- d_6) δ 45.9, 57.1, 73.9, 75.9, 76.3, 133.1, 134.3, 135.4, 137.3, 138.2, 145.3, 163.2, 177.4; mass (ES+) $m/z=230$ (M⁺+1); λ_{max} =222 nm; HR-EIMS Calcd for C₁₃H₁₄N₃O: 229.1215, found: 229.1218.

4.4.9. 6-Methyl-5-p-tolyl-5,6,7,8-tetrahydro-imidazo[1,2 a]**pyrimidin-2-one (13c).** Yield: 52% as a brown oil; R_f =0.5 (CHCl₃/MeOH, 90:10); v_{max} (neat) 1668 (CONH), 3357 (NH) cm⁻¹; ¹H NMR (200 MHz, CDCl₃) δ 0.83 (d, 3H, $J=6.6$ Hz, CH₃), 0.88 (d, 3H, $J=6.6$ Hz, CH₃), 2.36 (s, 6H, $2 \times \text{ArCH}_3$), 2.53–2.56 (m, 2H, $2 \times \text{CHCH}_2$), 3.03–3.15 $(m, 4H, 2 \times CH_2NH), 3.76$ (s, 2H, CH₂CO), 3.77 (s, 2H, CH₂CO), 4.44 (d, 2H, $J=5.2$ Hz, $2\times$ CHPh), 6.98 (d, 4H, $J=8.0$ Hz, 2×2 ArH), 7.23 (d, 4H, $J=8.0$ Hz, 2×2 ArH); ¹³C NMR (75 MHz, CDCl₃) δ 13.4, 14.6, 19.8, 29.6, 33.1, 39.5, 42.0, 51.6, 58.5, 61.5, 125.7, 126.1, 127.7, 128.2, 128.5, 132.0, 133.9, 137.0, 165.7, 182.9; mass (ES+) $m/z=244$ (M⁺+1); $\lambda_{\text{max}}=225 \text{ nm}$; HR-EIMS Calcd for $C_{14}H_{17}N_3O: 243.1372$, found: 243.1365.

4.4.10. 5-(2,4-Dichloro-phenyl)-6-methyl-5,6,7,8-tetrahydro-imidazo[1,2-a]pyrimidin-2-one (13g). Yield: 57% as a brown oil; $R_f=0.7$ (CHCl₃/MeOH, 90:10); v_{max} (neat) 1660 (CONH), 3350 (NH) cm^{-1} ; ¹H NMR (200 MHz, CDCl₃) δ 0.83 (d, 3H, J=6.4 Hz, CH₃), 0.88 (d, 3H, $J=6.4$ Hz, CH₃), 2.23–2.40 (m, 2H, 2×CH–CH₂), 2.49– 2.71 (m, 4H, $2 \times CH_2NH$), 3.67 (s, 2H, CH₂CO), 3.73 (s, 2H, CH₂CO), 4.07 (d, 1H, J=1.4 Hz, CHPh), 4.13 (d, 1H, J=1.4 Hz, CHPh), 7.03–7.09 (m, 2H, 2×ArH), 7.30–7.47

 $(m, 2H, 2 \times ArH), 7.53$ (s, 1H, ArH), 7.55 (s, 1H, ArH); ¹³C NMR (75 MHz, CDCl₃) δ 11.9, 13.2, 14.2, 15.1, 28.4, 29.3, 32.9, 34.7, 39.7, 42.5, 44.5, 51.5, 58.2, 61.1, 126.8, 127.0, 127.4, 127.6, 127.8, 128.2, 128.3, 133.6, 133.8, 134.6, 170.4, 180.7; mass (ES+) $m/z=298$ (M⁺+1); $\lambda_{\text{max}}=223$ nm; HR-EIMS Calcd for $C_{13}H_{13}Cl_2N_3O$: 297.0436, found: 297.0438.

4.4.11. 7-Benzyl-3-methyl-4-phenyl-3,4,6,7,8,9-hexahydro-pyrimido $[1,2-a]$ pyrimidin-2-one (24). Yield: 63% as colorless oil; t_R =19.3 min; ν_{max} (neat) 1688 (CONH), 3412 (NH) cm⁻¹; ¹H NMR (200 MHz, CDCl₃) δ 0.84– 0.97 (m, 3H, CH3), 1.05–1.33 (m, 3H, CH3), 2.19–2.42 $(m, 2H, 2 \times CH), 2.49 - 2.75$ $(m, 4H, 2 \times CH_2), 2.96 - 3.35$ $(m, 10H, 2\times 2CH_2 \text{ and } CH)$, 3.43–3.67 $(m, 2H, 2\times CH)$, 3.92–4.09 (m, 1H, CH), 4.11–4.29 (m, 1H, CH) 7.05–7.36 $(m, 20H, ArH), 10.61$ (br s, 2H, 2×NH); ^{13}C NMR (50 MHz, CDCl3) d 12.5, 33.3, 37.4, 43.5, 50.06, 51.38, 60.8, 66.3, 67.2, 68.7, 76.9, 77.6, 78.2, 127.0, 127.3, 128.5, 129.4, 138.5, 139.5, 157.6, 177.8; λ_{max} = 215 nm; mass $(ES+)$ $m/z=334$ (M^++1) ; HR-EIMS Calcd for $C_{21}H_{23}N_3O: 333.1841$, found: 333.1840.

4.4.12. 3-Methyl-7-(4-methyl-benzyl)-4-phenyl-3,4,6,7, 8,9-hexahydro-pyrimido[1,2-a]pyrimidin-2-one (25). Yield: 56% as colorless oil; $t_R=18.1$ min; v_{max} (neat) 1690 $(CONH)$, 3431 (NH) cm⁻¹; ¹H NMR (200 MHz, CDCl₃) δ 0.87–0.93 (m, 3H, CH₃), 1.24–1.29 (m, 3H, CH₃), 2.32 (s, 3H, CH3), 2.35 (s, 3H, CH3), 2.60–2.74 (m, 2H, $2 \times$ CH), 2.85–3.37 (m, 12H, 2×3 CH₂), 3.59–3.78 (m, 2H, 2-CH), 3.92–4.01 (m, 1H, CH), 4.05–4.13 (m, 1H, CH), 6.89–7.22 (m, 18H, 2×9ArH), 10.50 (br s, 2H, 2×NH); ¹³C NMR (50 MHz, CDCl₃) δ 12.5, 17.3, 17.7, 21.5, 33.4, 37.6, 39.4, 43.7, 50.2, 127.0, 128.2, 128.4, 129.2, 130.1, 133.9, 134.4, 136.4, 138.8, 157.8, 178.3; $\lambda_{\text{max}} = 224 \text{ nm}$; mass $(ES+)$ $m/z=348$ $(M^+ + 1)$; HR-EIMS Calcd for C₂₂H₂₅N₃O: 347.1998, found: 347.1999.

4.4.13. 3-Methyl-4-phenyl-7-(4-trifluoromethyl-benzyl)- 3,4,6,7,8,9-hexahydro-pyrimido[1,2-a]pyrimidin-2-one (26). Yield: 68% as colorless oil; t_R =21.1 min; v_{max} (neat) 1683 (CONH), 3409 (NH) cm^{-1} ; ¹H NMR (200 MHz, CDCl3) d 0.86–0.96 (m, 3H, CH3), 1.09–1.36 (m, 3H, CH₃), 2.03–2.19 (m, 2H, 2×CH), 2.58–2.79 (m, 4H, $2 \times CH_2$), 3.13–3.42 (m, 4H, $2 \times CH_2$), 3.72–4.21 (m, 6H, $2 \times CH_2$ and $2 \times CH$), 7.32–7.49 (m, 10H, ArH), 7.57–7.73 $(m, 8H, ArH)$ 10.32 (br s, 2H, 2×NH); ¹³C NMR (50 MHz, CDCl3) d 14.3, 15.5, 43.8, 44.8, 47.2, 51.6, 56.5, 61.9, 64.3, 66.3, 104.4, 114.7, 117.3, 126.6, 128.5, 129.5, 130.8, 132.0, 144.4, 145.1, 148.5, 173.0; $\lambda_{\text{max}} = 222 \text{ nm}$; mass $(ES+)$ m/z 402 $(M^+ + 1)$; HR-EIMS Calcd for $C_{22}H_{22}F_3N_3O: 401.1715$, found: 401.1718.

4.4.14. 7-(4-Chloro-benzyl)-3-methyl-4-(4-trifluoromethyl-phenyl)-3,4,6,7,8,9-hexahydro-pyrimido[1,2 a]pyrimidin-2-one (27). Yield: 67% as colorless oil; t_R =21.8 min; v_{max} (neat): 1686 (CONH), 3457 (NH) cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 0.85-0.97 (m, 3H, CH₃), 1.30–1.38 (m, 3H, CH₃), 2.21–2.46 (m, 2H, 2×CH), 2.53–2.80 (m, 6H, $2 \times CH_2$ and $2 \times 1H$ of CH₂), 2.83–3.29 $(m, 6H, 2 \times CH_2 \text{ and } 2 \times 1H \text{ of } CH_2)$, 3.60–3.72 $(m, 2H,$ 2-CH), 4.01–4.10 (m, 1H, CH), 4.15–4.24 (m, 1H, CH), 6.84–7.03 (m, 4H, ArH), 7.19–7.40 (m, 8H, ArH), 7.58–

7.65 (m, 4H, ArH), 10.54 (br s, 2H, $2\times$ NH); ¹³C NMR (50 MHz, CDCl3) d 14.4, 51.9, 61.4, 61.5, 111.0, 118.4, 121.8, 125.8, 125.9, 127.0, 128.4, 129.6, 129.9, 130.5, 131.2, 131.9, 136.8, 141.4, 143.2, 145.3, 166.2; $\lambda_{\text{max}} =$ 221 nm; mass (ES+) $m/z = 436$ (M⁺+1); HR-EIMS Calcd for C22H21ClF3N3O: 435.1325, found: 435.1321.

4.5. Typical procedure for the preparation of 8

To a stirred solution of $6(6.33 \text{ mmol}, 1.0 \text{ g})$ in THF (50 mL) was added K_2CO_3 (9.49 mmol, 1.31 g) followed by ethyl iodoacetate (7.59 mmol, 0.89 mL) and the reaction was allowed to continue for 1.5 h at room temperature. Thereafter, the reaction mixture was quenched with water (50 mL) and extracted with EtOAc $(3 \times 30 \text{ mL})$. The organic layers were combined, dried (Na_2SO_4) , and evaporated under vacuo to yield a residue. This crude product was purified via silica gel column chromatography using hexane/EtOAc (90:10, v/v) to afford 7 followed by hexane/EtOAc (80:20, v/v) to furnish 8.

4.5.1. [(3-Cyano-4-phenyl-but-3-enyl)-ethoxycarbonylamino]-acetic acid ethyl ester (7). Yield: 4% as colorless oil; R_f =0.8 (hexane/EtOAc; 80:20); v_{max} (neat) 1736 $(2 \times C\dot{O}_2Et)$, 2218 (CN) cm⁻¹; ¹H NMR (200 MHz, CDCl₃) δ 1.29 (t, 6H, J=7.1 Hz, 2×CH₂CH₃), 3.65 (s, 4H, $CH₂$), 3.77 (s, 2H, CH₂), 4.20 (q, 4H, J=7.1 Hz, $2 \times CH_2CH_3$), 7.23 (s, 1H, =CH), 7.42–7.44 (m, 3H, ArH), 7.78–7.80 (m, 2H, ArH); mass (ES+) $m/z = 331$ $(M^+ + 1)$; Anal. Calcd for C₁₈H₂₂N₂O₄: C, 65.44; H, 6.71; N, 8.48. Found: C, 65.42; H, 6.59; N, 8.66.

4.5.2. (3-Cyano-4-phenyl-but-3-enyl)-carbamic acid ethyl ester (8). Yield: 58% as colorless oil; R_f =0.7 (hexane/ EtOAc; 80:20); v_{max} (neat) 1738 (CO₂Et), 2213 (CN) cm⁻¹; ¹H NMR (200 MHz, CDCl₃) δ 1.30 (t, 3H, $J=7.2$ Hz, CH₂CH₃), 1.97 (br s, 1H, NH), 3.48 (s, 2H, CH₂), 3.63 (d, 2H, $J=1.2$ Hz, CH₂), 4.23 (q, 2H, $J=7.2$ Hz, CH_2CH_3), 7.13 (s, 1H, $=CH$), 7.42–7.46 (m, 3H, ArH), 7.76–7.78 (m, 2H, ArH); 13C NMR (75 MHz, CDCl3) d 12.9, 53.5, 57.1, 57.2, 60.0, 106, 107.1, 117.1, 127.7, 129.4, 131.7, 144.0, 144.9, 169.0; mass (ES+) $mlz=$ 245 (M⁺+1); Anal. Calcd for C₁₄H₁₆N₂O₂: C, 68.83; H, 6.60; N, 11.47. Found: C, 69.17; H, 6.47; N, 11.66.

4.6. General procedure for the of preparation of 11c,g

To a solution of appropriate acetate^{[6](#page-7-0)} from $5c, g$ (2.5 mmol) in a THF/H₂O system (10 mL, 1:1, v/v) was added DABCO (0.42 g, 3.76 mmol) at room temperature and the mixture was stirred for 10 min. Simultaneously, in another flask to the solution of glycine methyl ester hydrochloride (0.47 g, 3.75 mmol) in $H₂O$ (5 mL) was added Et₃N (0.52 mL, 5.0 mmol) and the mixture was stirred for 10 min. This mixture was then added dropwise to the flask containing the acetate and the reaction was continued at room temperature for 1.5 h. The THF was removed from the reaction mixture and the residue was extracted with EtOAc $(2\times20$ mL) and water (25 mL). The combined organic layers were dried $(Na₂SO₄)$ and concentrated in vacuo to afford the crude product. Purification of the crude product via column chromatography over silica gel with hexane/EtOAc (85:15, v/v) furnished the pure products.

4.6.1. (2-Cyano-1-p-tolyl-allylamino)-acetic acid methyl ester (11c). Yield: 76% as colorless oil; $R_f=0.6$ (hexane/ EtOAc; 85:15); v_{max} (neat) 1737 (CO₂Me), 2226 (CN) cm⁻¹; ¹H NMR (200 MHz, CDCl₃) δ 1.88 (br s, 1H, NH), 2.35 (s, 3H, CH₃), 3.36 (d, 2H, $J=2.5$ Hz, CH₂CO), 3.71 $(s, 3H, OCH₃), 4.41$ $(s, 1H, CH), 5.98$ $(s, 1H, =CH₂),$ 6.09 (s, 1H, $=CH_2$), 7.17 (d, 2H, $J=8.0$ Hz, ArH), 7.28 (d, 2H, J=8.0 Hz, ArH); ¹³C NMR (75 MHz, CDCl₃) δ 19.9, 46.7, 50.7, 63.4, 116.2, 125.0, 126.0, 128.4, 128.8, 134.3, 137.2, 171.2; mass (ES+) $mlz=245$ (M⁺+1); Anal. Calcd for $C_{14}H_{16}N_2O_2$: C, 68.83; H, 6.60; N, 11.47. Found: C, 69.11; H, 6.66; N, 11.55.

4.6.2. [2-Cyano-1-(2,4-dichloro-phenyl)-allylamino]-acetic acid methyl ester (11g). Yield: 81% as colorless oil; $R_f = 0.8$ (hexane/EtOAc; 85:15); v_{max} (neat) 1732 $(CO₂Me)$, 2216 (CN) cm⁻¹; ¹H NMR (200 MHz, CDCl₃) δ 3.38 (d, J=5.6 Hz, 2H, CH₂CO), 3.74 (s, 3H, OCH₃), 4.94 (s, 1H, CH), 6.05 (s, 1H, =CH₂), 6.13 (s, 1H, $=CH₂$), 7.31–7.42 (m, 2H, ArH), 7.61–7.66 (d, 1H, $J=8.4$ Hz, ArH); ¹³C NMR (75 MHz, CDCl₃) δ 46.6, 50.8, 62.9, 115.8, 124.4, 127.5, 127.9, 129.4, 133.2, 135.8, 171.1; mass (ES+) $m/z=299$ (M⁺+1), 301 (M⁺+3); Anal. Calcd for $C_{13}H_{12}Cl_2N_2O_2$: C, 52.19; H, 4.04; N, 9.36. Found: C, 51.87; H, 3.95; N, 9.29.

4.7. General procedure for the preparation of 16–19

To a stirred solution of appropriate acetate from 14 and 15 (5.2 mmol) in THF/H₂O $(5 \text{ mL}, 1:1, v/v)$ system was added DABCO (7.8 mmol, 0.87 g,) at room temperature and the reaction mixture was allowed to stir for 5 min. This was followed by dropwise addition of a solution of appropriate allyl amine from $6a-c$ (5.2 mmol) in THF (5 mL) and the reaction was allowed to proceed till completion (as analyzed by TLC, ca. 1 h). Thereafter excess THF was evaporated in vacuo and the residue was extracted with EtOAc $(3\times30 \text{ mL})$ and water 50 mL. The organic layers were combined, dried (Na_2SO_4) , and evaporated in vacuo to obtain a residue, which was purified via silica gel column chromatography using hexane/EtOAc (85:15, v/v) to yield the pure product.

4.7.1. 2-[(2-Cyano-3-phenyl-allylamino)-phenyl-methyl] acrylic acid ethyl ester (16). Yield: 68% as colorless oil; R_f =0.5 (hexane/EtOAc; 85:15); ν_{max} (neat) 1713 (CO₂Et), 2212 (CN), 3340 (NH) cm⁻¹; ¹H NMR (200 MHz, CDCl₃) δ 1.24 (t, 3H, J=7.2 Hz, CH₂CH₃), 3.53 (s, 2H, CH₂NH), 4.17 (q, 2H, J=7.2 Hz, CH₂CH₃), 4.79 (s, 1H, CHNH), 6.01 (s, 1H, $=CH_2$), 6.41 (s, 1H, $=CH_2$), 7.05 (s, 1H,]CH), 7.27–7.46 (m, 8H, ArH), 7.74–7.77 (m, 2H, ArH); ¹³C NMR (75 MHz, CDCl₃) δ 12.9, 13.1, 43.3, 50.3, 52.2, 59.8, 108.9, 117.2, 124.5, 126.5, 127.3, 127.4, 127.6, 128.3, 129.0, 132.0, 133.7, 139.4, 140.5, 142.8, 165.0, 166.7; mass (ES+) $m/z = 347$ (M⁺+1). Anal. Calcd for C22H22N2O2: C, 76.28; H, 6.40; N, 8.09. Found: C, 76.41; H, 6.20; N, 7.92.

4.7.2. 2-[(2-Cyano-3-p-tolyl-allylamino)-phenyl-methyl] acrylic acid ethyl ester (17). Yield: 73% as colorless oil; R_f =0.6 (hexane/EtOAc; 85:15); ν_{max} (neat) 1712 (CO₂Et), 2217 (CN), 3399 (NH) cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 1.21 (t, 3H, J=7.2 Hz, CH₂CH₃), 2.38 (s, 3H, ArCH₃), 3.47 (s, 2H, CH₂NH), 4.14 (q, 2H, $J=7.2$ Hz, CH₂CH₃), 4.76 (s, 1H, CHNH), 6.01 (s, 1H, $=CH_2$), 6.39 (s, 1H, $=CH₂$), 6.97 (s, 1H, $=CH$), 7.19–7.42 (m, 7H, ArH), 7.64 (d, 2H, J=8.0 Hz, ArH); ¹³C NMR (75 MHz, DMSO- d_6) d 12.8, 13.0, 50.4, 59.8, 108.6, 117.0, 124.1, 124.2, 124.6, 125.3, 126.7, 127.5, 127.6, 127.9, 128.2, 128.5, 128.6, 129.1, 131.8, 139.8, 143.0, 143.7, 164.5; mass (ES+) $m/z=$ 361 (M⁺+1); Anal. Calcd for C₂₃H₂₄N₂O₂: C, 76.64; H, 6.71; N, 7.77. Found: C, 76.68; H, 6.50; N, 7.66.

4.7.3. 2-[(2-Cyano-3-phenyl-allylamino)-(4-trifluoromethyl-phenyl)-methyl]-acrylic acid ethyl ester (18). Yield: 89% as colorless oil; $R_f=0.8$ (hexane/EtOAc; 85:15); v_{max} (neat) 1714 (CO₂Et), 2214 (CN), 3343 (NH) cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 1.26 (t, 3H, $J=7.2$ Hz, CH₂CH₃), 3.54 (s, 2H, CH₂NH), 4.16 (q, 2H, $J=7.2$ Hz, CH_2CH_3), 4.84 (s, 1H, CHNH), 6.04 (s, 1H, $=CH_2$), 6.46 (s, 1H, $=CH_2$), 7.04 (s, 1H, $=CH$), 7.41– 7.51 (m, 3H, ArH), 7.56–7.63 (m, 4H, ArH), 7.72–7.77 (m, 2H, ArH); ¹³C NMR (75 MHz, DMSO- d_6) δ 12.8, 50.3, 59.8, 59.9, 109.3, 116.8, 124.3, 125.4, 126.7, 127.9, 128.8, 130.3, 135.1, 139.8, 141.6, 143.6, 164.6; mass (ES+) $m/z = 415$ (M⁺+1); Anal. Calcd for C₂₃H₂₁F₃N₂O₂: C, 66.66; H, 5.11; N, 6.76. Found: C, 66.50; H, 4.88; N, 6.92.

4.7.4. 2-[[3-(4-Chloro-phenyl)-2-cyano-allylamino]-(4 trifluoromethyl-phenyl)-methyl]-acrylic acid ethyl ester (19). Yield: 78% as colorless oil; v_{max} (neat) 1720 (CO_2Et) , 2214 (CN), 3400 (NH) cm⁻¹; ¹H NMR $(300 \text{ MHz}, \text{CDCl}_3)$ δ 1.27 (t, 3H, J=7.2 Hz, CH₂CH₃), 3.53 (d, 2H, J=2.1 Hz, CH₂NH), 4.20 (q, 2H, J=7.5 Hz, CH_2CH_3), 4.82 (s, 1H, CHNH), 6.01 (s, 1H, $=CH_2$), 6.45 $(s, 1H, =CH_2)$, 7.00 $(s, 1H, =CH)$, 7.37 (d, 2H, J=8.4 Hz, ArH), 7.47–7.70 (m, 4H, ArH), 7.68 (d, 2H, J=8.4 Hz, ArH); mass (ES+) m/z =449 (M⁺+1); Anal. Calcd for $C_{23}H_{20}CIF_3N_2O_2$: C, 61.54; H, 4.49; N, 6.24. Found: C, 61.68; H, 4.50; N, 6.56.

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